

**Supporting Information for**

**Engineering chlorophyll, bacteriochlorophyll and carotenoid  
biosynthetic pathways in *Escherichia coli***

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**Table S1. Genes used to assemble (B)Chl and carotenoid biosynthetic pathways in *E. coli*.**

Gene	Locus	Organism	Annotation
<i>chlI</i>	slr1030	<i>Synechocystis</i> sp. PCC 6803	I subunit of magnesium chelatase
<i>chlD</i>	slr1777	<i>Synechocystis</i> sp. PCC 6803	D subunit of magnesium chelatase
<i>chlH</i>	slr1055	<i>Synechocystis</i> sp. PCC 6803	H subunit of magnesium chelatase
<i>gun4</i>	slI0558	<i>Synechocystis</i> sp. PCC 6803	porphyrin-binding protein that enhances magnesium chelatase
<i>chlM</i>	slr0525	<i>Synechocystis</i> sp. PCC 6803	magnesium-protoporphyrin IX methyltransferase
<i>bciB</i>	slr1923	<i>Synechocystis</i> sp. PCC 6803	ferredoxin-dependent 8-vinyl reductase
<i>chlP</i>	slI1091	<i>Synechocystis</i> sp. PCC 6803	geranylgeranyl reductase
<i>chlG</i>	slr0056	<i>Synechocystis</i> sp. PCC 6803	chlorophyll <i>a</i> synthase
<i>acsF</i>	rge_33550	<i>Rubrivivax gelatinosus</i> IL144	O <sub>2</sub> -dependent magnesium-protoporphyrin IX monomethyl ester cyclase
<i>crtE</i>	rge_33730	<i>Rubrivivax gelatinosus</i> IL144	geranylgeranyl pyrophosphate synthase
<i>bchN</i>	rsp_0285	<i>Rhodobacter sphaeroides</i> 2.4.1	N subunit of dark-operative protochlorophyllide oxidoreductase
<i>bchB</i>	rsp_0286	<i>Rhodobacter sphaeroides</i> 2.4.1	B subunit of dark-operative protochlorophyllide oxidoreductase
<i>bchL</i>	rsp_0288	<i>Rhodobacter sphaeroides</i> 2.4.1	L subunit of dark-operative protochlorophyllide oxidoreductase
<i>bchC</i>	rsp_0263	<i>Rhodobacter sphaeroides</i> 2.4.1	3-hydroxyethyl bacteriochlorophyllide dehydrogenase
<i>bchX</i>	rsp_0262	<i>Rhodobacter sphaeroides</i> 2.4.1	X subunit of chlorophyllide oxidoreductase
<i>bchY</i>	rsp_0261	<i>Rhodobacter sphaeroides</i> 2.4.1	Y subunit of chlorophyllide oxidoreductase
<i>bchZ</i>	rsp_0260	<i>Rhodobacter sphaeroides</i> 2.4.1	Z subunit of chlorophyllide oxidoreductase
<i>bchF</i>	rsp_0284	<i>Rhodobacter sphaeroides</i> 2.4.1	3-vinyl bacteriochlorophyllide hydratase
<i>bchG</i>	rsp_0279	<i>Rhodobacter sphaeroides</i> 2.4.1	bacteriochlorophyll <i>a</i> synthase
<i>crt<sup>RS</sup></i>	rsp_0271	<i>Rhodobacter sphaeroides</i> 2.4.1	3-step phytoene desaturase
<i>crt<sup>BRS</sup></i>	rsp_0270	<i>Rhodobacter sphaeroides</i> 2.4.1	15- <i>cis</i> -phytoene synthase
<i>dxs</i>	b0420	<i>Escherichia coli</i>	1-deoxy-D-xylulose-5-phosphate synthase
<i>crt<sup>YPa</sup></i>	n/a	<i>Pantoea agglomerans</i>	lycopene cyclase
<i>crt<sup>IPa</sup></i>	n/a	<i>Pantoea agglomerans</i>	4-step phytoene desaturase
<i>crt<sup>BPa</sup></i>	n/a	<i>Pantoea agglomerans</i>	15- <i>cis</i> -phytoene synthase

**Table S2. Strains and plasmids described in this study.**

Strain/Plasmid	Characteristics	Source
<b><u>E. coli</u></b>		
JM109	Cloning strain for plasmid construction	Promega
C43(DE3)	Expression strain for <i>in vivo</i> assay and assembly of (B)Chl and carotenoid biosynthesis pathways	Ref. 1
<b><u>Synechocystis</u></b>		
WT	sp. PCC 6803, glucose tolerant	R. Sobotka <sup>†</sup>
<b><u>Rba. sphaeroides</u></b>		
WT	2.4.1	S. Kaplan <sup>‡</sup>
$\Delta bchP$	Unmarked deletion of the <i>bchP</i> gene in WT	Ref. 2
$\Delta crtC$	Unmarked deletion of the <i>crtC</i> gene in WT	Ref. 3
<b><u>Plasmid</u></b>		
pET3a	Expression vector carrying T7 promoter, Amp <sup>R</sup>	Novagen
pACYCDuet1	Expression vector with two multiple cloning sites (MCS) both preceded by a T7lac promoter, Cm <sup>R</sup>	Novagen
pCDFDuet1	Expression vector with two MCS both preceded by a T7lac promoter, Sm <sup>R</sup>	Novagen
pCOLADuet1	Expression vector with two MCS both preceded by a T7lac promoter, Km <sup>R</sup>	Novagen
pAC-BETA	Contains <i>Pantoea agglomerans crtE</i> , <i>crtY</i> , <i>crtI</i> , and <i>crtB</i> genes with their native promoters and enables <i>E. coli</i> to produce $\beta$ -carotene, Cm <sup>R</sup>	Ref. 4
pET3a- <i>dvr</i>	<i>Synechocystis dvr</i> gene (internal <i>SpeI</i> site removed) cloned into <i>NdeI/SpeI</i> sites of a modified pET3a with an <i>SpeI</i> site added immediately upstream of the <i>BamHI</i> site (same applies for all pET3a constructs described in this study), Amp <sup>R</sup>	Ref. 5
pET3a- <i>chlG</i>	<i>Synechocystis chlG</i> gene cloned into <i>NdeI/SpeI</i> sites of pET3a, Amp <sup>R</sup>	Ref. 5
pET3a- <i>bchCXYZ</i>	<i>Rba. sphaeroides bchCXYZ</i> genes cloned into <i>NdeI/SpeI</i> sites of pET3a, Amp <sup>R</sup>	This study
pET3a- <i>bchF</i>	<i>Rba. sphaeroides bchF</i> genes cloned into <i>NdeI/SpeI</i> sites of pET3a, Amp <sup>R</sup>	This study
pET3a- <i>bchG</i>	<i>Rba. sphaeroides bchG</i> genes cloned into <i>NdeI/SpeI</i> sites of pET3a, Amp <sup>R</sup>	This study
pET3a- <i>bchNBL</i>	<i>Rba. sphaeroides bchNBL</i> genes (internal <i>NdeI</i> and <i>SpeI</i> sites removed) cloned into <i>NdeI/SpeI</i> sites of pET3a, Amp <sup>R</sup>	This study
pET3a- <i>crtE</i>	<i>Rvi. gelatinosus crtE</i> gene cloned into <i>NdeI/SpeI</i> sites of pET3a, Amp <sup>R</sup>	This study
pET3a- <i>crtYIB</i>	<i>Pantoea agglomerans crtYIB</i> gene fragment amplified from pAC-BETA and cloned into <i>NdeI/SpeI</i> sites of pET3a, Amp <sup>R</sup>	This study
pET3a- <i>bchCXYZFG</i>	Link and lock cloning, <i>bchCXYZ-bchF-bchG</i> cloned into pET3a, Amp <sup>R</sup>	This study
IA	Link and lock cloning, <i>chlI-chlD-chlH-gun4-chlM-acsF</i> cloned into pET3a, Amp <sup>R</sup>	Sci. Adv.
P1-1	Link and lock cloning, <i>Synechocystis chlG</i> gene cloned downstream of the <i>acsF</i> gene of IA, Amp <sup>R</sup>	This study
P1-2	Link and lock cloning, <i>Rba. sphaeroides bchNBL</i> genes cloned downstream of the <i>acsF</i> gene of IA, Amp <sup>R</sup>	This study
BoP	The BoWSCP-His <sub>10</sub> coding sequence and <i>Synechocystis chlP</i> gene cloned into the <i>NcoI/HindIII</i> sites and <i>NdeI/XhoI</i> sites of pACYCDuet1, Cm <sup>R</sup>	Ref. 5
pCDFDuet1*- <i>dvr</i>	The <i>XbaI-HindIII</i> region containing the <i>lacI</i> gene and T7lac promoter 1 of pCDFDuet1 replaced with the T7 promoter- <i>XbaI-HindIII</i> fragment of pET3a- <i>dvr</i> , Sm <sup>R</sup>	This study
pCDFDuet1*- <i>dvr-2-chlP</i>	The <i>HindIII/XhoI</i> fragment containing the <i>chlP</i> gene cut from BoP and cloned into pCDFDuet1*- <i>dvr</i> , Sm <sup>R</sup>	This study
P2-1	Link and lock cloning, <i>Rba. sphaeroides bchNBL</i> genes cloned downstream of the <i>dvr</i> gene of pCDFDuet1*- <i>dvr-2-chlP</i> , Sm <sup>R</sup>	This study
P2-2	The <i>XbaI/HindIII</i> fragment containing <i>bchCXYZFG</i> genes cut from pET3a- <i>bchCXYZFG</i> and cloned into pCDFDuet1*- <i>dvr</i> to replace the <i>dvr</i> gene, Sm <sup>R</sup>	This study

<b>P2-3</b>	The <i>XbaI/HindIII</i> fragment containing <i>bchCXYZFG</i> genes cut from P2-2 and cloned into pCDFDuet1*- <i>dvr-2-chIP</i> to replace the <i>dvr</i> gene, Sm <sup>R</sup>	This study
<b>P3-1</b>	The DE plasmid reported in Sci. Adv. paper with <i>E. coli dxs</i> and <i>Rvi. gelatinosus crtE</i> genes cloned into <i>NcoI/HindIII</i> sites and <i>NdeI/XhoI</i> sites of pCOLADuet1, renamed as plasmid P3-1 in this study for clarity, Km <sup>R</sup>	Ref. 5
pCOLADuet1- <i>dxs</i>	<i>E. coli dxs</i> gene cloned at the <i>NcoI/HindIII</i> sites of pCOLADuet1	This study
pCOLADuet1*- <i>crtE</i>	<i>Rvi. gelatinosus crtE</i> gene with the adjacent <i>SpeI-HindIII</i> region of pET3a amplified from pET3a- <i>crtE</i> and cloned into <i>NcoI/HindIII</i> sites of pCOLADuet1, Km <sup>R</sup>	This study
<b>P3-2</b>	<i>Rvi. gelatinosus crtE</i> gene and <i>Rba. sphaeroides crtIB</i> genes with a 42-bp RBS-containing sequence (with the last T changed to A) from pET3a added between <i>crtE</i> and <i>crtIB</i> cloned into <i>NdeI/KpnI</i> sites of pCOLADuet1- <i>dxs</i> , Km <sup>R</sup>	This study
<b>P3-3</b>	Link and lock cloning, <i>Pantoea agglomerans crtYIB</i> gene fragment cloned downstream of the <i>crtE</i> gene of pCOLADuet1*- <i>crtE</i> , Km <sup>R</sup>	This study

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**Table S3. Oligonucleotide primers used in this study.**

<b>Primer</b>	<b>Sequence (5'-3')</b>
bchCFNdeI	GGAACATATGGTGAGAACGACCGCCGTCAT
bchZRSpeI	GGAAACTAGTTCATTGGTTCTCTCCCTTCCTCT
bchFFNdeI	GGAACATATGCAGCCCACGTCCCCCGC
bchFRSpeI	GGAAACTAGTTCATTGCGCGGCTCCATGTC
bchGFNdeI	GGAACATATGAGTGTCAATCTATCCTTACA
bchGRSpeI	GGAAACTAGTTCACGGCAGCACCTCCAGCC
bchNFNdeI	TCTCATATGAGCCTTGACCTTCCGCC
bchNBremoveNdeIF	CTGACGCTGTGGACATACGAAGGCCCGCCCCATGTG
bchNBremoveNdeIR	CACATGGGGCGGGCCTTCGTATGTCCACAGCGTCAG
bchNBremoveHindIIIF	GCAGATCTGCCGCAAGCTCGCCAGGCCCATGGAGCG
bchNBremoveHindIIIR	CGCTCCATGGGCCTGGCGAGCTTGCGGCAGATCTGC
bchBLfusionF	AGCTCATTATGCACGGTGAGCGGACGGGACGGCAAG
bchBLfusionR	CTTGCCGTCCCGTCCGCTCACCGTGCATAATGAGCT
bchLRSpeI	TCTACTAGTTC AATCGAAACCCAGCAACTC
crtEFNdeI	TCTCATATGAACACGATGACTCGCATCGA
crtERSpeI	TCTACTAGTTC AAGCGGTCTGGGTCGGAG
crtYPaFNdeI	GGCCATATGAGGGATCTGATTTTAGTCGG
crtBPaRSpeI	GGCACTAGTCTAAACGGGACGCTGCCAAAGA
CDFDuetLLF	GCGAAGCTTGCGGCCGATAATGC
CDFDuetLLR	GCGTCTAGAGGGAAACCGTTGTGGTCTCCCTATAGTGAGTCGTATTAGCGGTTCAAGTAGAAAAGATCA
crtEFNcoI	GGCCCATGGGTAACACGATGACTCGCATCGAACA
pET3aRHindIII	GGCAAGCTTTAATGCGGTAGTTTATCAC
crtERBSR	TTGTATATCTCCTTCTTAAAGTTAAACAAAATTTATTTCTAGTTCAAGCGGTCTGGGTCGGAG
RBSertIRsF	ACTAGAAATAATTTTGTTTAACTTTAAGAAGGAGATATACAAATGCCCTCGATCTCGCCCGC
crtBRsRKpnI	GGAAGGTACCCTAGATCGGGTTGGCCCGGTT