## **Supporting Information**

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Fig. S1. Amino acid sequence alignments of known AcsF proteins. Sequences are those from *Rvi. gelatinosus* (AcsF), *Synechocystis* (Cycl), *C. reinhardtii* (CRD1), *A. thaliana* (CHL27), and *Rba. sphaeroides* (Rsp\_0294; abbreviated as 0294). Conserved, highly similar, and similar residues are highlighted in black, dark gray, and light gray, respectively. The putative diiron center ligands are marked by red diamonds.



**Fig. S2.** Genetic knockouts and replacements in *Rvi. gelatinosus.* (*A*) Depiction of the deletion of *bchE* (*Left*), confirmed by colony PCR (*Right*). (*B*) Depiction of deletion of *acsF*, and subsequent integration of foreign genes at the *acsF* locus, under control of the native promoter (*Upper*), confirmed by colony PCR (*Lower*). The regions subjected to genetic manipulation are depicted in proportion to the scale bar. ORFs are represented as colored filled rectangles, within which the arrow indicates the direction of transcription. Crt, carotenoid biosynthesis; RC&LHC, reaction center and light-harvesting complexes.



Fig. S3. Deletion of rsp\_6110 in Rba. sphaeroides. Diagram depicting deletion of rsp\_6110 (Left), and confirmation by colony PCR (Right).



**Fig. S4.** Construction and phenotypic analysis of *Rvi. gelatinosus* mutant expressing *bciE* and *acsF* from *Rba. sphaeroides*. (A) Diagram depicting integration of *bciE* and *acsF* from *Rba. sphaeroides* in place of the native *acsF* in *Rvi. gelatinosus* (*Upper*), and confirmation by colony PCR (*Lower*). (B) HPLC analysis of pigments extracted from *Rvi. gelatinosus* strains, extracted from the same number of cells of each strain except for the  $\Delta bchE$  strain, which had a much greater BChl *a* content compared with the other strains. (*Inset*) Retention times and Soret/Q<sub>y</sub> maxima of peaks were used to identify BChl *a*.klj.



**Fig. S5.** Current status of known components of the oxygen-dependent cyclase.  $AcsF^{\alpha}$ ,  $AcsF^{Anox}$ , and  $AcsF^{Ox}$  represent AcsF proteins from Alphaproteobacteria, anoxygenic phototrophs, respectively.  $e^-$  denotes the electron donor to the diiron center of AcsF.

Table S1. Distribution of *acsF* and *bchE* genes among sequenced phototrophic Proteobacteria, along with the presence of orthologs of rsp\_6110

			Presence	:e/abse	nce of:
Organism	GenBank no.	Group	6110	acsF	bchE
Acidiphilium multivorum AIU301	AP012035	α1 <i>,</i> ΑΑΡ	•	•	0
Phaeospirillum molischianum DSM 120	ZP_09875400	α1, PNB	Х	х	0
Rhodospirillum centenum SW	CP000613	α1, PNB	•	•	0
Rhodospirillum rubrum ATCC 11170	CP000230	α1, PNB	Х	х	0
Rhodospirillum photometricum DSM 122	YP_005416037	α1, PNB	х	х	0
Ahrensia sp. strain R2A130	NZ_AEEB01000017	α2 <i>,</i> ΑΑΡ	•	•	х
Agrobacterium albertimagni AOL15	ALJF0000000	α2, ΑΑΡ	•	•	0
Hoeflea phototrophica DFL43	NZ_ABIA02000022	α2 <i>,</i> ΑΑΡ	•	•	х
Labrenzia alexandrii DFL11	NZ_EQ973123.1	α2 <i>,</i> ΑΑΡ	•	•	0
Methylobacterium sp. strain 4-46	CP000943	α2 <i>,</i> ΑΑΡ	•	•	0
Methylobacterium radiotolerans	CP001001	α2 <i>,</i> ΑΑΡ	•	•	х
Methylobacterium populi BJ001	YP_001927978	α2 <i>,</i> ΑΑΡ	•	•	х
Methylobacterium extorquens AM1	YP_002966142	<b>α2, AAP</b>	•	•	х
Methylocella silvestris BL2	CP001280	α2, AAP	•	•	х
Bradyrhizobium sp. strain BTAi1	CP000494	α2, PNB	•	•	٠
Bradyrhizobium sp. strain ORS278	CU234118	α2, PNB	•	•	•
Rhodomicrobium vannielii ATCC 17100	NC 014664	α2, PNB	x	x	0
Rhodopseudomonas palustris	Multiple	α2, PNB	•	•	0
Dinoroseobacter shibae DFL12	CP000830	<b>α3. AAP</b>	•	•	0
Jannaschia sp. strain CCS1	CP000264	α3. ΑΑΡ	•	•	X
Loktanella vestfoldensis SKA53	AAMS0100000	α3. ΔΔΡ			x
Rosenbacter denitrificans Och 114	CP000362	α3 ΔΔΡ			0
Roseobacter litoralis Och 149	ARIG000000				0
Rosephacter sn strain AzwK-3h					•
Rosephacter sp. strain A2WK-30					x
Rosenvarius sn strain TM1025		α3, ΑΑΓ α2 ΔΔD			<u>^</u>
Roseovarius sp. strain 1101035		α3, ΑΑΓ α3 ΔΔΡ			0
Rhodohacter canculatus SR 1002		as DND	Y	v l	•
Rhodobacter sphaeroides	Multinle	as phip			-
Rhodobacter spinerolaes	7P 05842911	as PNR			•
					- V
<i>Erythrobacter</i> sp. strain NAP1	AAIVIWUUUUUUU0000	α4, AAP	•	•	X
Citromicrobium bathyomarinum JL354	ZP_06861151	α4, AAP	•	•	0
Sphingomonas spp.	Multiple	α4, ΑΑΡ	•	•	X
Brevundimonas subvibrioides ATCC 15264	CP002102	α4, ΑΑΡ	•	•	X
Rubrivivax gelatinosus IL-114	NC_017075	β <i>,</i> PNB	Х	٠	0
Rubrivivax benzoatilyticus JA2	NZ_AEWG01000000	β <i>,</i> PNB	Х	•	0
Methyloversatilis universalis FAM5	ZP_08506871	β <i>,</i> PNB	Х	٠	Х
Limnohabitans sp. strain Rim28	ALKN0000000	β <i>,</i> PNB	Х	•	Х
Limnohabitans sp. strain Rim47	ALKO0000000	β <i>,</i> PNB	Х	٠	x
Allochromatium vinosum DSM 180	NC_013851	γ, PSB	х	х	0
Ectothiorhodospira sp. strain PHS-1	NZ_AGBG01000002	γ, PSB	Х	х	0
Halorhodospira halophila SL1	CP000544	γ, PSB	Х	х	•
Marichromatium purpuratum 984	NZ_AFWU01000001	γ, PSB	х	х	0
Thiocapsa marina 5811	NZ_AFWV01000003	γ, PSB	Х	х	•0
Thiocystis violascens DSM198	AGFC00000000	γ <i>,</i> PSB	х	х	0
Thioflavicoccus mobilis 8321	NC_019940	γ <i>,</i> PSB	х	х	0
Thiorhodococcus drewsii AZ1	NZ_AFWT01000007	γ, PSB	х	х	0
Thiorhodospira sibirica ATCC 700588	 AGFD01000016	γ, PSB	х	х	0
Congregibacter litoralis KT71	AAOA01000014	γ, AAP	х	•	•
gamma proteobacterium NOR5-3	ZP 05125815	v, AAP	X	•	•
Luminiphilus syltensis NOR51-B	NZ DS999411	v. AAP	X	•	х
marine aamma proteobacterium HTCC2080	) NZ DS999405	v. AAP	x	•	X
gamma proteobacterium HIMB55	ZP 09691978	v, AAP	X	•	X

Modified from (26).  $\bullet$ , gene present in PGC;  $\bigcirc$ , gene present outside PGC; X, gene absent. The red box indicates an identical pattern of presence/absence of orthologs of rsp\_6110 and *acsF* among Alphaproteobacteria. AAP, aerobic anoxygenic phototroph; PNB, purple nonsulfur bacterium; PSB, purple sulfur bacterium.

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## Table S2. Strains and plasmids described in this study

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Strain/plasmid	Genotype/characteristics	Source
E. coli		
JM109	Cloning strain for plasmid constructs	Promega
S17-1	Conjugation strain for pK18mobsacB constructs	(48)
Rvi. gelatinosus		
WT	IL144	S. Nagashima <sup>3</sup>
∆bchE	Unmarked deletion mutant of <i>bchE</i> in WT	This study
$\Delta bchE\Delta acsF$	Unmarked deletion mutant of <i>acsF</i> in $\Delta bchE$	This study
$\Delta bchE\Delta acsF::acsF^{Rs}$	acs $F^{Rs}$ replacement of acsF in $\Delta bchE$	This study
$\Delta bchE\Delta acsF::bciE-acsF^{Rs}$	acsF replaced with rsp 6110-acsF <sup>Rs</sup> in $\Delta bchE$	This study
$\Delta bchE\Delta acsF::cycl$	cvcl replacement of acsF in $\Delta bchE$	This study
$\Delta bchE\Delta acsF::cvcl-vcf54$	$cvcl-vcf54$ replacement of acsF in $\Delta bchE$	This study
Synechocystis		, <b>,</b>
ŴT	sp. PCC6803	R. Sobotka <sup>†</sup>
acsF <sup>Rg+</sup>	acs $F^{Rg}$ and $Km^{R}$ replacement of psbAll in WT	This study
$acsF^{Rg+}\Delta cvcl$	$Cm^{R}$ replacement of cvcl in acs $F^{R_{g+}}$	This study
$acsF^{Rg+} \Delta cvcl \Delta vcf54$	Zeo <sup>R</sup> replacement of central portion of vcf54 in acs $F^{Rg+}\Delta cvcl$	This study
∆vcf54	Zeo <sup>R</sup> replacement of central portion of vcf54 in WT	(22)
Rba. sphaeroides	······································	
wt	2.4.1	S. Kaplan <sup>‡</sup>
$\Delta bchE\Delta ccoP$	Unmarked deletion mutant of <i>bchE</i> and <i>ccoP</i> in WT	(15)
$\Delta bchE\Delta ccoP\Delta acsF$	Unmarked deletion mutant of <i>acsF</i> in $\Delta bchE\Delta ccoP$	(15)
$\Delta bchE\Delta ccoP\Delta 6110$	Unmarked deletion mutant of rsp 6110 in $\Delta bchE\Delta ccoP$	This study
Plasmids		,
pK18mobsacB	Allelic exchange vector, <i>Km<sup>R</sup></i>	J. Armitage <sup>§</sup>
pK18∆ <i>bchE<sup>Rg</sup></i>	Upstream-Ndel-downstream of bchE <sup>Rg</sup> cloned into BamHI/HindIII sites of pK18mobsacB	This study
pK18∆ <i>acsF<sup>Rg</sup></i>	Upstream-Ndel-downstream of acsF <sup>Rg</sup> cloned into BamHI/HindIII sites of pK18mobsacB	This study
pK18∆6110	Upstream-downstream of rsp_6110 cloned into Xbal/HindIII sites of pK18mobsacB	This study
pK18[ <i>acsF<sup>Rs</sup></i> ]	acs $F^{Rs}$ cloned into the Ndel site of pK18 $\Delta$ acs $F^{Rg}$	This study
pK18[6110-acsF <sup>Rs</sup> ]	rsp_6110- <i>acsF<sup>Rs</sup></i> cloned into the <i>Ndel</i> site of pK18∆ <i>acsF<sup>Rg</sup></i>	This study
pK18[ <i>cycl</i> ]	cycl cloned into the Ndel site of pK18 $\Delta acsF^{Rg}$	This study
pK18[cycl-ycf54]	<i>cycI-ycf54</i> cloned into the <i>NdeI</i> site of pK18∆ <i>acsF<sup>Rg</sup></i>	This study
pPD-FLAG	Cloning site, Km <sup>R</sup> , flanked by psbAll upstream and downstream regions, Amp <sup>R</sup>	(21)
pPD[ <i>acsF<sup>Rg</sup></i> ]	acsF <sup>Rg</sup> cloned into Ndel/Bg/II sites of pPD-FLAG	This study
pBBRBB-Ppuf <sub>843-1200</sub>	Expression vector carrying the 843–1,200 region of <i>puf</i> promoter of <i>Rba. sphaeroides</i> , Km <sup>R</sup>	(27)
pBB[6110]	rsp_6110 cloned into the BglII/NotI sites of pBBRBB-Ppuf <sub>843-1200</sub>	This study

\*Research Institute for Photosynthetic Hydrogen Production, Kanagawa University, Yokohama, Japan. <sup>†</sup>Institute of Microbiology, Department of Phototrophic Microorganisms, Třeboň, Czech Republic. <sup>‡</sup>Department of Microbiology and Molecular Genetics, University of Texas Medical School, Austin, TX. <sup>§</sup>Department of Biochemistry, University of Oxford, Oxford, United Kingdom.

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## Table S3. Primers used in this study

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Primer	Sequence (5′-3′)		
6110UpF	GCTCTAGAGGAGCTGATCCCGCCCTTCC		
6110UpR	GGAGAGCCCTCCGGCCGGCGCGTTCATGGGGGGTTCCCTTCTCTTGG		
6110DownF	CCAAGAGAAGGGAACCCCCATGAACGCGCCGGCCGGAGGGCTCTCC		
6110DownR	GCAAGCTTCCCAGGTTCACCGCCACGCC		
6110CheckF	GCCCCGGAGCGACAAGGAC		
6110CheckR	GTATTTCTTGGCCTTGGTCAGG		
6110F_Ndel	GAGTCTCATATGGGTCTGTTCACGAAACAAGCG		
6110F_Bglll	GGCAGATCTATGGGTCTGTTCACGAAACAAGCGGAA		
6110R_Notl	TCTGCGGCCGCTCACAGCGTCACCTGCTCGGAGAA		
0294F_Ndel	CCAGTACATATGTGAACGCGCCGGCCGGAGG		
0294R_Ndel	CCAGTACATATGTCAATAGCTCGGCTCCAGTCGG		
45840UpF	CTAGGTCAAGTAGGATCCTCATGCCGGCGGCGATCATG		
45840UpR	CTAGGTCAAGTACATATGGGAAACGGCTCCTCGCGATTC		
45840DownF	CTAGGTCAAGTACATATGCGACGGCTGGGTCACGATGC		
45840DownR	CTAGGTCAAGTAAAGCTTTGCCGGTGTAGAAGTCGCACGC		
45840CheckF	TAGCCGCCGACCATGCCGA		
45840CheckR	GCGGTGCACCAGCACCGTGA		
33550UpF	GAGTCTGGATCCCTGCATGAGCGACAACGCGTC		
33550UpR	GAGTCTCATATGGAGGGTCTCCGTGGTGTGTCA		
33550DownF	GAGTCTCATATGAAGCGAGGACAGGATGCTGAGC		
33550DownR	GAGTCTAAGCTTGGAACTCCTCGCTCAGGTTGCG		
33550CheckF	GAACGTTTGCCGGACACGGT		
33550CheckR	ACGAGGTACTTCAGGTGCTCC		
33550F_Ndel	GAGTCTCATATGCTCGCGACCCCGACGATCG		
33550R_BamHI	GAGTCTGGATCCTCACCATGCCGGGGGCCATG		
1214UpF	GCCGATCCGGTTAACCTAGGCA		
1214UpR	ATATCCAGTGATTTTTTTTCTCCATAGAGTTGTTTAAAATAGTTTCC		
1214UpCmF	GGAAACTATTTTAAACAACTCTATGGAGAAAAAAATCACTGGATAT		
1214DownCmR	GGTGATCCAGCGGAAGACAACCTTACGCCCCGCCCTGC		
1214DownF	GCAGGGCGGGGCGTAAGGTTGTCTTCCGCTGGATCACC		
1214DownR	TGGAGTTGTTGGGAGAGTTCGGTC		
1214F_Ndel	GGAATTCCATATGGTTAATACCCTCGAAAAGCCCG		
1214R_Ndel	GGAATTCCATATGTTAGCGCACAGCTCCAGCCA		
1214RBS1780F	GTTGGCTGGAGCTGTGCGCTAATATAGGAGCTTGGATTGTGGAAAGTTGGGCATTGACGA		
1214RBS1780R	TCGTCAATGCCCAACTTTCCACAATCCAAGCTCCTATATTAGCGCACAGCTCCAGCCAAC		
1780F	GTGGAAAGTTGGGCATTGACG		
1780R	CTAATCCAGGGATGCAAGGGG		
1780R_Ndel	GAGTCTCATATGCTAATCCAGGGATGCAAGGGG		